

CYNANCHOSIDE, A HIGHLY OXYGENATED IRIDOID GLUCOSIDE FROM *MACFADYENA CYNANCHOIDES*

CARLO BONINI, ENRICO DAVINI, CARLO IAVARONE and CORRADO TROGOLO

Centro di Studio per la Chimica delle Sostanze Organiche Naturali del C.N.R., Roma; Istituto di Chimica Organica dell'Università di Roma, Piazzale Aldo Moro, 5-00185-Roma, Italy

(Revised received 6 October 1980)

Key Word Index—*Macfadyena cynanchoides*; Bignoniaceae; iridoid glucoside; cynanchoside.

Abstract—The elucidation of the structure and stereochemistry of cynanchoside, a new highly oxygenated iridoid glucoside isolated from *Macfadyena cynanchoides* (Bignoniaceae), has been accomplished using mainly ^1H and ^{13}C NMR spectral data and further confirmed by simple chemical transformations.

INTRODUCTION

Re-investigation of the iridoid fraction present in the ethanolic extract of leaves of *Macfadyena cynanchoides* (Bignoniaceae), collected in the autumn, revealed the presence, in addition to macfadyenoside (**1**) [1] of at least five other compounds with a possible iridoid structure. In this paper we report the structure of the most polar compound, which was present in small amounts and which we have named cynanchoside (**2**).

RESULTS AND DISCUSSION

Compound **2** was an amorphous compound with molecular formula $\text{C}_{15}\text{H}_{24}\text{O}_{12}$ and $[\alpha]_{\text{D}} -126^\circ$, which gave a brown colour with the vanillin reagent. Its UV (204 nm, $\log \epsilon = 3.6$) and IR (1670 cm^{-1}) absorptions indicated the presence of a non-conjugated iridoid enol-ether system. By enzymatic hydrolysis with β -glucosidase, **2** gave D-glucose (1 mol) thus permitting the identification of the compound as a β -D-glucopyranoside.

The low R_f value of **2** was typical of highly polar iridoids. Its colour reaction and the co-occurrence of **1** in the plant as the major iridoid component suggested at first the possible identity of **2** with calycinoside (**3**) [2] (5-O- β -D-glucosylmacfadyenoside, R_f 0.04, brown reaction with vanillin). However, unlike **1** and **3**, compound **2** did not give a positive Ross test [3] for the oxirane function.

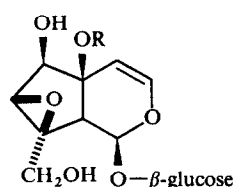
The ^1H NMR spectrum of **2** (Table 1), which showed the typical doublet for the anomeric proton of a β -glucopyranosyl moiety (δ 4.76, $J = 7.5\text{ Hz}$), closely resembled that of 7 α -hydroxyharpagide (**4**) [4] except for the presence of an AB system for 10- CH_2OH ($\delta_{\text{A}} = 3.81$, $\delta_{\text{B}} = 3.57$, $J_{\text{AB}} = 13.0\text{ Hz}$) instead of the sharp singlet at δ 1.05 of the methyl group (C-10) in **4**.

Assuming the same absolute configurations at C-1, C-5 and C-9 as found in other iridoid glucosides [5], the small coupling constant $J_{1,9}$ (ca 0 Hz, dihedral angle ca 90°) provided evidence for an axial position of the β -D-glucopyranosyl moiety at C-1.

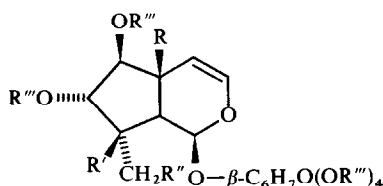
Acetylation of **2** under mild conditions afforded the heptaacetate **5** whose ^1H NMR spectrum exhibited, in comparison with that of **2**, paramagnetic shifts in agreement with the presence in the aglycone moiety of one primary ($\Delta\delta \approx 0.5$) and two secondary ($\Delta\delta \approx 1.2$) alcohol functions.

By further acetylation, **5** was completely transformed into the octaacetate **6** owing to the acetylation of one of the two tertiary OH functions (OH-8). In fact **6** showed a residual hydroxyl absorption (OH-5) in the IR spectrum, in agreement with the different reactivity of tertiary OH functions at C-5 and C-8 towards acetylation, which represents a characteristic feature of iridoid glucosides [5].

The structure 7 α ,10-dihydroxyharpagide suggested for **2** was strengthened by these data and conclusively



- 1 $\text{R} = \text{H}$
3 $\text{R} = \beta\text{-glucose}$



- 2 $\text{R} = \text{R}' = \text{R}'' = \text{OH}$, $\text{R}''' = \text{H}$
4 $\text{R} = \text{R}' = \text{OH}$, $\text{R}'' = \text{R}''' = \text{H}$
5 $\text{R} = \text{R}' = \text{OH}$, $\text{R}'' = \text{OAc}$, $\text{R}''' = \text{Ac}$
6 $\text{R} = \text{OH}$, $\text{R}' = \text{R}'' = \text{OAc}$, $\text{R}''' = \text{Ac}$
7 $\text{R} = \text{R}''' = \text{H}$, $\text{R}' = \text{R}'' = \text{OH}$

Table 1. ^1H NMR shift assignments (90 MHz, δ values)

Compound	H-1	H-3	H-4	H-6	H-7	H-9	2H-10 3H-10	AcO-	Isopr.
2 (D_2O)	5.86 <i>s</i>	6.42 <i>d</i> $J_{3,4} = 6.0$	5.26 <i>d</i>	3.97 <i>d</i> $J_{6,7} = 9.0$	3.65 <i>d</i>	2.45 <i>s</i>	3.81 } 3.57 } AB $J_{AB} = 13.0$		
4* (D_2O)	5.61 <i>s</i>	6.30 <i>d</i> $J_{3,4} = 6.5$	5.15 <i>d</i>	3.81 <i>d</i> $J_{6,7} = 9.0$	3.50 <i>d</i>	2.37 <i>s</i>	1.05 <i>s</i>		
5 (CDCl_3)	5.62 <i>s</i>	6.22 <i>d</i> $J_{3,4} = 6.5$	5.44 <i>d</i>	4.8–5.3	4.8–5.3	2.88 <i>s</i>	4.30 } 4.06 } AB $J_{AB} = 12.0$	1.8–2.1	
6 (CDCl_3)	5.60 <i>s</i>	6.28 <i>d</i> $J_{3,4} = 6.3$	4.8–5.3	5.65† <i>d</i> $J_{6,7} = 9.0$	4.9–5.6	3.54 <i>s</i>	4.03 $\frac{1}{2}$ AB $J_{AB} = 12.0$	1.9–2.2	
9 (D_2O)	5.71 <i>s</i>	6.42 <i>d</i> $J_{3,4} = 6.5$	5.21 <i>d</i>	3.97‡ <i>d</i> $J_{6,7} = 9.0$	3.25‡ <i>d</i>	2.31 <i>s</i>	3.93 <i>s</i>		1.33 1.24
10 (D_2O)	5.66 <i>s</i>	6.52 <i>d</i> $J_{3,4} = 6.0$	5.26 <i>d</i>	4.05§ <i>d</i> $J_{6,7} = 6.0$	4.23§ <i>d</i>	2.66 <i>s</i>	4.06 AB $J_{AB} = 12.0$		1.50 (3 H) 1.39 (9 H)
11 (CDCl_3)	5.61 <i>s</i>	6.21‡ <i>d</i> $J_{3,4} = 6.3$	5.39‡ <i>bd</i>	5.65‡ <i>d</i> $J_{6,7} = 8.5$	4.51‡ <i>d</i>	2.76 <i>s</i>	3.97 <i>bs</i>	2.12, 2.08 2.05, 2.01	1.38 1.27
12 (CDCl_3)	5.57 <i>s</i>	6.34‡ <i>d</i> $J_{3,4} = 6.3$	4.98‡ <i>d</i>	3.93‡ <i>d</i> $J_{6,7} = 6.0$	5.44‡ <i>d</i>	2.79 <i>s</i>	3.98 <i>bs</i>	2.14, 2.09 2.05, 2.00 1.98	1.49, 1.36 1.31, 1.26

* These data are relative to a new spectrum we registered to complete those previously reported.

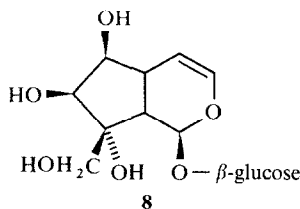
† The high field line of this doublet is overlapped by the H-1 signal.

‡ Assignment checked by spin decoupling experiments.

§ These assignments could be reversed.

confirmed by the ^{13}C NMR spectrum of **2** (Table 2) whose resonances were practically coincident with those of **4** apart from the presence in **2** of a triplet (SFORD) at 62.11 ppm ($10\text{-CH}_2\text{OH}$) instead of the corresponding quartet arising from the methyl group in **4**. This coincidence points to the same stereochemistry for all chiral centres in **2** and **4**. In particular, the *trans*-1,2-diol arrangement of the hydroxyl functions at C-6 and C-7 of **2** was well supported by the rather low field value [7] of their resonances, as found also in **4**.

The stereochemistry of the C-8 centre was inferred to be identical in **2** and **4** taking into account the differing effects of an $8\alpha\text{-OH}$ and an $8\beta\text{-OH}$ substituent upon the resonance of C-9. In fact, it has been observed [7] in the pair of C-8 isomers 10-*des*-cynnamoylglobularinin (**7**) and 10-*des*-cynnamoylglobularinin (**8**) that the β -configuration of OH-8 causes the deshielding of C-9 (δ 48.04) as compared to its α -counterpart (δ 43.70). Analogous effects have been noticed [8] in another pair epimeric at the quaternary C-8, gardenoside (β OH-8, C-9 δ 52.40) and monotropein methyl ester (α OH-8, C-9 δ 45.42).



Since **7** may be considered a 5-deoxy derivative of **2**, we have calculated from suitable pairs of iridoids, differing only by the presence of the OH-5, that the deshielding effect of the OH-5 on the C-9 resonance is approximately 7 ppm (7.21 ppm in hastatoside [9] and verbenalin [9]; 7.54 ppm in macfadyenoside [9] and catalpol (unpublished results). The subtraction of this shift increment from the resonance of C-9 (δ 56.16) of **2** gives a resonance value in better agreement with that of C-9 of **7** (δ 48.04) than with that of C-9 of **8** (δ 43.70), indicating an identical stereochemistry of the C-8 centre in **2**, **4** and **7**.

Conclusive chemical evidence for the relative orientation of the secondary and tertiary hydroxyl groups in the aglycone moiety of **2** was obtained by reaction of **2** with acetonedimethylketal- SnCl_2 , which afforded the *O*-*iso*-propylidene derivatives **9** and **10**.

The ^1H NMR spectrum of **9** compared with that of **2** shows the signals of only one *O*-*iso*-propylidene group (methyl singlets at δ 1.33 and 1.24) and small shift changes for H-6, H-7, H-9 and 10-CH_2 which therefore do not represent useful clues for placing the *iso*-propylidene unit.

A comparison of the ^{13}C NMR spectrum of **9** with that of **2** reveals a large downfield shift for the C-8 of **9** ($\Delta\delta = +8.20$), analogous to the α -effect caused by the acetylation of a tertiary OH function, and smaller and different effects on C-10 ($\Delta\delta = +1.57$), C-6 ($\Delta\delta = -4.01$), C-9 ($\Delta\delta = -2.44$) and C-7 ($\Delta\delta = -1.46$).

The acetylation of **9** led to the hexaacetate **11** whose ^1H NMR spectrum showed notable downfield shifts for H-6 and H-7 ($\Delta\delta$ 1.68 and 1.26, respectively) showing

Table 2. ^{13}C NMR chemical shifts*

Compound	2 (D ₂ O)	4† (D ₂ O)	5‡ (CDCl ₃)	7§ (CD ₃ OD)	8§ (CD ₃ OD/DMSO)	9 (D ₂ O)	10 (D ₂ O)
C-1	91.69 <i>d</i>	92.01 <i>d</i>	91.31 <i>d</i>	93.34	95.16	91.32 <i>d</i>	91.41 <i>d</i>
C-3	140.55 <i>d</i>	139.96 <i>d</i>	139.74 <i>d</i>	140.39	141.61	140.14 <i>d</i>	143.54 <i>d</i>
C-4	109.28 <i>d</i>	109.48 <i>d</i>	110.06 <i>d</i>	106.54	105.27	109.38 <i>d</i>	105.96 <i>d</i>
C-5	64.95 <i>s</i>	64.17 <i>s</i>	64.45 <i>s</i>	37.32	37.16	64.15 <i>s</i>	75.79 <i>s</i>
C-6	82.53 <i>d</i>	82.64 <i>d</i>	83.98 <i>d</i>	83.14	78.34	78.52 <i>d</i>	87.71 <i>d</i>
C-7	78.90 <i>d</i>	78.92 <i>d</i>	78.32 <i>d</i>	86.42	79.34	77.44 <i>d</i>	80.19 <i>d</i>
C-8	76.27 <i>s</i>	74.91 <i>s</i>	75.41 <i>s</i>	80.33	81.03	84.47 <i>s</i>	88.87 <i>s</i>
C-9	56.16 <i>d</i>	56.36 <i>d</i>	56.83 <i>d</i>	48.04	43.70	53.72 <i>d</i>	52.26 <i>d</i>
C-10	62.11 <i>t</i>	16.69 <i>q</i>	63.47 <i>t</i>	64.29	66.37	63.68 <i>t</i>	63.88 <i>t</i>
Me-C							115.33 <i>s</i>
						110.17 <i>s</i>	110.26 <i>s</i>
							28.25 <i>q</i>
							27.72 <i>q</i>
Me-C						26.67 <i>q</i>	26.38 <i>q</i>
						25.70 <i>q</i>	25.88 <i>q</i>
C-1'	98.86	98.74	95.89			98.92	99.03
C-2'	73.35	73.36	71.30			73.35	73.25
C-3'	76.27	76.28	72.24			76.28	76.16
C-4'	70.60	70.61	68.37			70.59	70.59
C-5'	77.05	77.06	72.24			76.86	76.85
C-6'	61.63	61.63	61.71			61.43	61.44

* The spectra were recorded at 20 MHz. Chemical shifts in ppm from TMS (dioxane (67.4 ppm) was used as internal standard).

† From ref. [9].

‡ Additional signals from acetoxy groups at 172.55, 170.68, 170.01, 169.33 ppm (C=O), 21.00 and 20.69 ppm (Me).

§ From ref. [7].

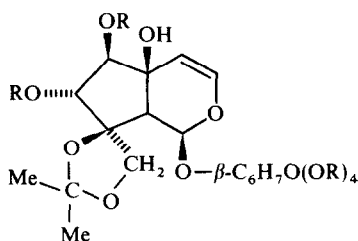
acetylation at these centres. This clearly confirmed that the ketal ring of **9** must be located at C-8 and C-10 and that this position, new in iridoids, is evidently preferred to that at C-5 and C-6.

The ^1H NMR spectrum of the second *O*-iso-propylidene derivative **10** showed that it contained two *iso*-propylidene units (methyl singlets at δ 1.50 (3 H) and 1.39 (9 H) respectively). The comparison of the ^{13}C NMR spectrum of **10** with the spectra of **9** and **2** respectively, revealed significant deshielding values for the carbons involved in the ketal functions: C-5 ($\Delta\delta = +11.64$; $+10.84$), C-6 ($\Delta\delta = +9.19$; $+5.18$), C-8 ($\Delta\delta = +4.40$; $+12.60$) and C-10 ($\Delta\delta = +0.20$; $+1.77$). As for the remaining carbons, the upfield shift of C-4 ($\Delta\delta = -3.42$; -3.32) was notable while the C-5' ($\Delta\delta = +3.40$; $+2.99$) and C-7 ($\Delta\delta = +2.75$; $+1.29$) resonances were slightly

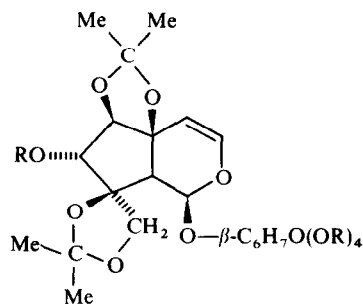
deshielded. As expected, the acetylation of **10** led to the pentaacetate (peracetate) **12** whose ^1H NMR spectrum showed four distinct methyl singlets and a downfield shift for H-7 ($\Delta\delta$ 1.21). All these data established a 5,6-8,10-*bis*-*O*-*iso*-propylidene derivative structure for compound **10**.

The ^{13}C NMR spectra of these *O*-*iso*-propylidene derivatives of iridoid glucosides, which are the first to be reported, allow it to be concluded that carbons bearing primary and tertiary hydroxyl functions are slightly (*ca* 1–2 ppm) and strongly (*ca* 8–12 ppm) deshielded, respectively, when involved in the formation of *O*-*iso*-propylidene units while the effects on carbons bearing secondary hydroxyl functions seem to be less distinct.

The formation of **10** proved a *cis*-relationship between the OH groups at C-5 and C-6, and consequently both are β -orientated. The fact that no 6,7-*iso*-propylidene



9 R = H
11 R = Ac



10 R = H
12 R = Ac

derivative was formed [10], combined with the ^{13}C NMR data, confirms the *trans*-relationship between the 6-OH and 7-OH groups and thus the α -orientation of the latter.

The present results show the value of ^{13}C NMR spectroscopy as an aid to solve configurational problems in the iridoid glucosides. The structure and configuration now demonstrated for cynanchoside (**2**, 7 α ,10-dihydroxyharpagide) is suggested to arise from **1** by cleavage of the epoxide ring.

EXPERIMENTAL

CC was on Si gel, 70–230 mesh (Merck) and cellulose CF 11 (Whatman). Si gel SIF₂₅₄ (Erba) and cellulose (Merck) plates were used for TLC. Paper chromatograms (PC) were on Schleicher and Schüll No. 2043 b Mgl paper. Spray reagents: 2 N H₂SO₄, heating at 120° (Si gel plates), vanillin (vanillin 1 g, conc HCl 2 ml, MeOH 100 ml) and 3,5-dinitrosalicylic acid (3,5-dinitrosalicylic acid 0.5 g, NaOH 4 g, H₂O 100 ml), heating at 100° (cellulose plates and PC).

Isolation of iridoid fraction. *Macfadyena cynanchoides* was harvested in autumn in the Botanical Garden, University of Rome. A reference specimen (A5-1) has been deposited at the Botanical Institute Herbarium, University of Rome. Fresh aerial part (4 kg) was extracted twice with 90% EtOH (10 l. each) at room temp. for 3 days. PC (*n*-BuOH–HOAc–H₂O, 63:10:27) visualized with vanillin, showed 6 spots with the following *R_f* values: 0.42 (*A*, brown), 0.33 (*B*, pink) 0.29 (*C*, pink), 0.16 (*D*, grey-violet), 0.11 (macfadyenoside (**1**), brown), 0.05 (cynanchoside (**2**), brown). The ethanolic extracts were concd to an aq. suspension which was continuously extracted with petrol, bp 40–70° (2 l.). Decolorizing charcoal (1 kg) was added and the aq. suspension stratified on a Gooch funnel (20 cm dia) containing a layer of Si gel (100 g). Mono- and di-saccharides were removed with H₂O (12 l.) and 5% EtOH (8 l.). Compounds **1** and **2** (fraction I, 5 g) with 10% EtOH (8 l.); **1**, **2** and *D* (fraction II, 3 g) with 20% EtOH (7 l.) and finally a mixture of **1**, *A*, *B*, *C* and *D* with 50% EtOH (5 l.) (fraction III, 2.5 g) and 80% EtOH (4 l.) (fraction IV, 0.5 g).

Isolation of cynanchoside (2). Fractions I and II (8 g), chromatographed on cellulose (250 g) in *n*-BuOH satd with H₂O afforded the following fractions: (a) *C* and *D* (0.5 g), (b) **1** (3 g), (c) **1** and **2** (1 g). Fraction (c) was rechromatographed on Si gel eluted with Me₂CO–H₂O (9:1) and then on Si gel eluted with EtOAc–MeOH (7:3), to give pure **2** (400 mg) as an amorphous powder; $[\alpha]_D^{25} -126^\circ$ (MeOH; *c* 1.0); UV $\lambda_{\text{max}}^{\text{MeOH}}$ nm (log *ε*): 204 (3.6); IR $\nu_{\text{max}}^{\text{KBr}}$ cm⁻¹: 3400, 2940, 1670, 1075, 1025. (Found: C, 45.28; H, 6.25. C₁₅H₂₄O₁₂ requires: C, 45.45; H, 6.10%).

Heptaacetate (5) and octaacetate (6) from 2. Compound **2** (100 mg) dissolved in dry pyridine (0.3 ml) was treated with Ac₂O (0.6 ml) for 1 hr at room temp. MeOH was added and after 20 min the soln was evapd and the residue, chromatographed on Si gel in Et₂O–EtOAc (19:1), gave the heptaacetate **5** (60 mg) as an amorphous powder. (Found: C, 50.20; H, 5.67. C₂₉H₃₈O₁₉

requires: C, 50.43; H, 5.55%). Compound **5** (60 mg), acetylated for 3 hr as described above for **2**, gave a residue which was chromatographed on Si gel in Et₂O–EtOAc (19:1) to give the octaacetate **6** (40 mg) as an amorphous powder. $[\alpha]_D^{25} -141^\circ$ (MeOH; *c* 0.7). (Found: C, 50.63; H, 5.63. C₃₁H₄₀O₂₀ requires: C, 50.82; H, 5.50%).

8,10-O-Iso-propylidene cynanchoside (9) and 5,6-8,10-bis-O-iso-propylidene cynanchoside (10). Compound **2** (260 mg) was treated with a 15% soln of SnCl₂ in dry Me₂CO (4 ml) adding acetonedimethylketal (0.1 ml). The suspension was stirred at room temp. for 1.5 hr then poured into cold, satd NaHCO₃ soln. The resulting suspension was centrifuged and the residue washed twice with Me₂CO–H₂O (1:1, 10 ml). The collected solns showed on TLC (*n*-BuOH–MeOH–H₂O, 7:1:3) the presence of two major compounds with higher *R_f* than **2** and were evapd to dryness to give an amorphous residue which, chromatographed on Si gel with *n*-BuOH satd with H₂O, gave **10** (56 mg) and **9** (42 mg) as amorphous powders.

Hexaacetate (11) from 9. Compound **9** (42 mg) was treated with pyridine (0.5 ml) and Ac₂O (1 ml) for 2 hr at room temp. The mixture, worked up as previously described for **5**, afforded an amorphous residue which, on TLC in Et₂O, showed only the presence of **11**. The residue chromatographed on Si gel in Et₂O gave **11** (28 mg) as an amorphous powder. (Found: C, 52.09; H, 5.99. C₃₀H₄₀O₁₈ requires: C, 52.32; H, 5.86%).

Pentaacetate 12 from 10. Compound **10** (56 mg), acetylated as described for **5**, gave an amorphous residue which on TLC in Et₂O–hexane (4:1) showed only the presence of **12**. The residue chromatographed on Si gel in Et₂O–hexane (4:1) afforded **12** (30 mg) as an amorphous powder. (Found: C, 53.98; H, 6.26. C₃₁H₄₂O₁₇ requires: C, 54.22; H, 6.17%).

REFERENCES

1. Bianco, A., Guiso, M., Iavarone, C. and Trogolo, C. (1974) *Gazz. Chim. Ital.* **104**, 731.
2. Bianco, A., Guiso, M., Iavarone, C., Poccia, L. and Trogolo, C. (1979) *Gazz. Chim. Ital.* **109**, 561.
3. Ross, W. C. J. (1950) *J. Chem. Soc.* 2257.
4. Scarpati, M. L., Guiso, M. and Esposito, P. (1968) *Gazz. Chim. Ital.* **98**, 177.
5. Bobbitt, J. M. and Segebarth, K. P. (1969) *Cyclopentanoid Terpene Derivatives* (Taylor, W. I. and Battersby, A. R. eds.) p. 1. M. Dekker, New York.
6. Steyn, R. and Sable, H. Z. (1971) *Tetrahedron* **27**, 4429.
7. Chaudhuri, R. K. and Sticher, O. (1979) *Tetrahedron Letters* 3149.
8. Chaudhuri, R. K., Afifi-Yazar, F. U. and Sticher, O. (1979) *Helv. Chim. Acta* **62**, 1603.
9. Bianco, A., Caciola, P., Guiso, M., Iavarone, C. and Trogolo, C. (1981) *Gazz. Chim. Ital.* (in press).
10. Bianco, A., Guiso, M., Iavarone, C. and Trogolo, C. (1975) *Gazz. Chim. Ital.* **105**, 185.